



Microbial communities in root zones of
Pennisetum setaceum (African fountain grass) at a
heavy metal contaminated site, Kabwe, Zambia

Chiba, A.^{1*}, Mweetwa, A.² and Uchida, Y.³

¹ Research Faculty of Veterinary Medicine, Hokkaido University, Japan

² Department of Soil Science, University of Zambia, Zambia

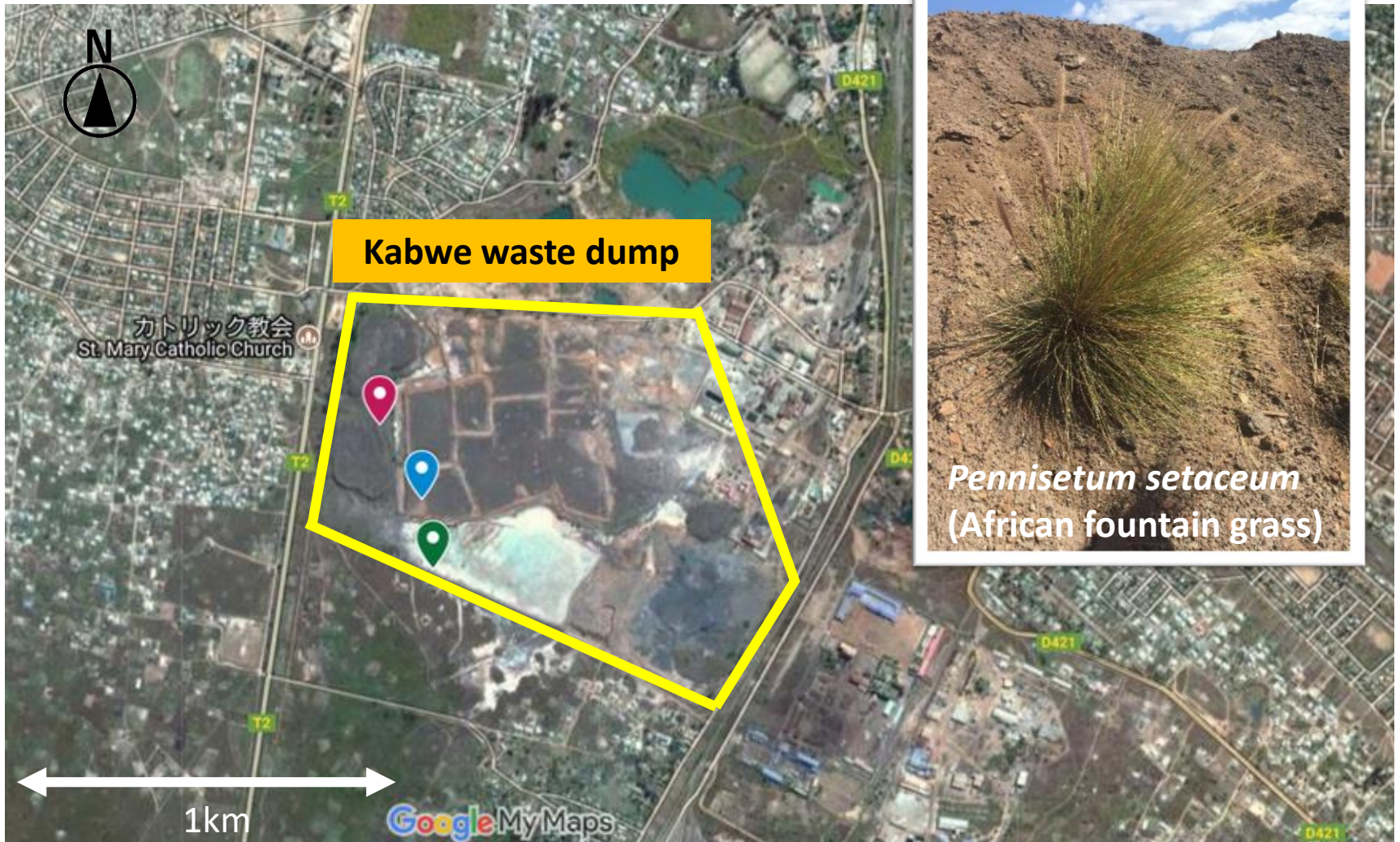
³ Research Faculty of Agriculture, Hokkaido University, Japan

Email: chiba@chem.agr.hokudai.ac.jp

Brief view of this study

1. Microbes in the root zone of African fountain grass in Kabwe, Zambia
2. A new portable sequencer for microbial DNA analysis

A waste dump at the center of Kabwe city



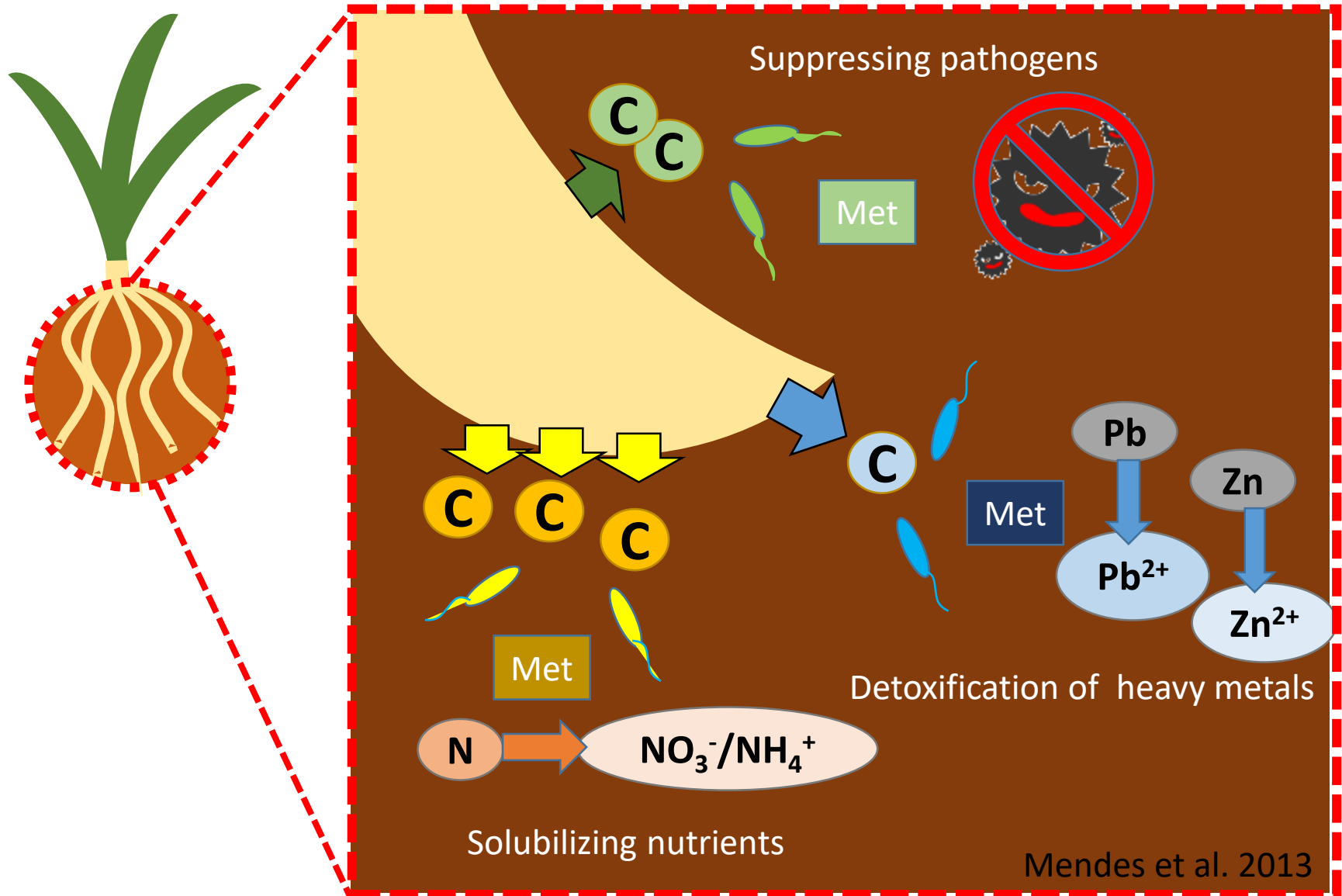
Why can they grow in such a harsh environment?

(1) Morphology



The unique characteristics protect African fountain grass from several stresses (e.g. heat and drought).

(2) Microbes in a root zone



Previous technique



- Isolation of single microbial species
- 1% of soil microbes can be isolated

New technique



- Multiple microbial species can be identified at the same time
- Non-culturable microbes are included

DNA level studies for microbial community

DNA samples
from any environments



Iontorrent
PGM



Reference database

Escherichia coli
GGTCATTAATACTA

Obtained sequence data

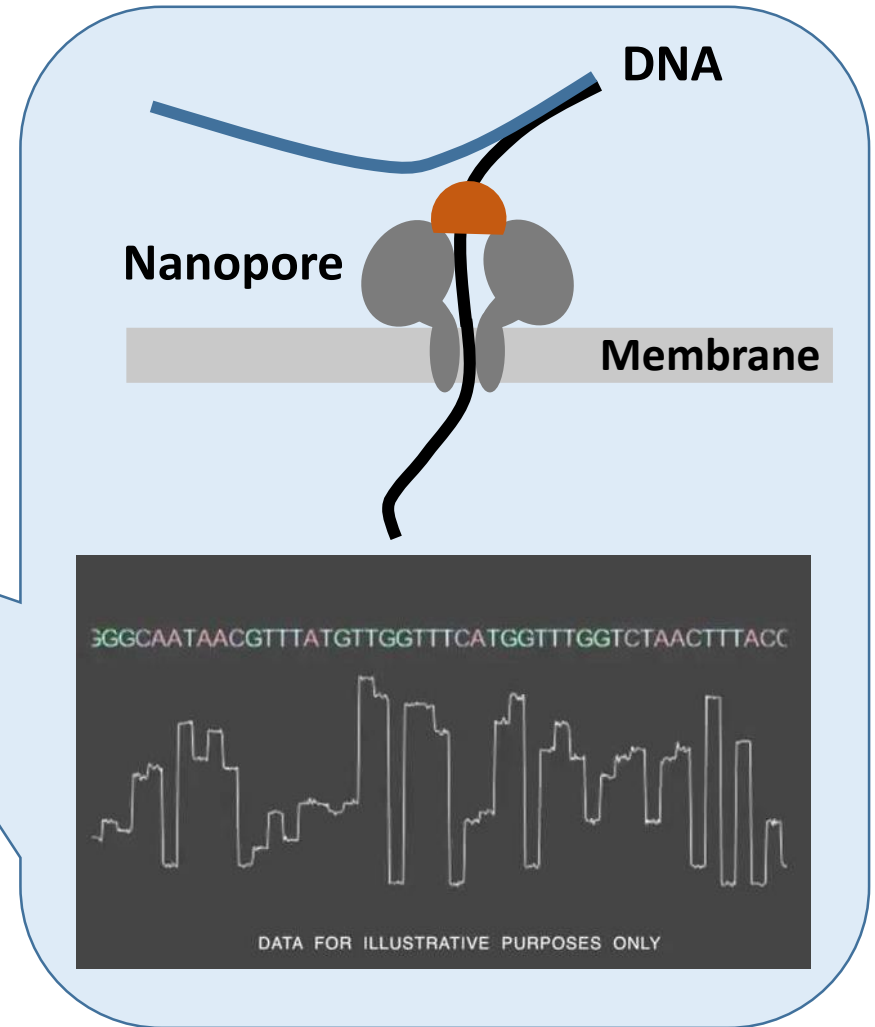
GGTCATTAATACTA
AGCCTTCCCATATAG
CGTTTCCTTCCATAA
ACTTCATATAGTTGA
GGCCTTATAGCCCAG

- High running cost
- Clean and well-equipped environments

New sequencing device “MinION” (Oxford Nanopore)



- USB-like device
- Reusable



We have to evaluate the usability of MinION sequencer for analyzing microbial community in Zambia

The purposes of the study

- 1. To identify microbes in the root zone of African fountain grass in Kabwe, Zambia**
- 2. To evaluate the usability of MinION sequencer for analyzing microbial community**

Sampling in June, 2017

- African fountain grass
- Different colored soils

Site A

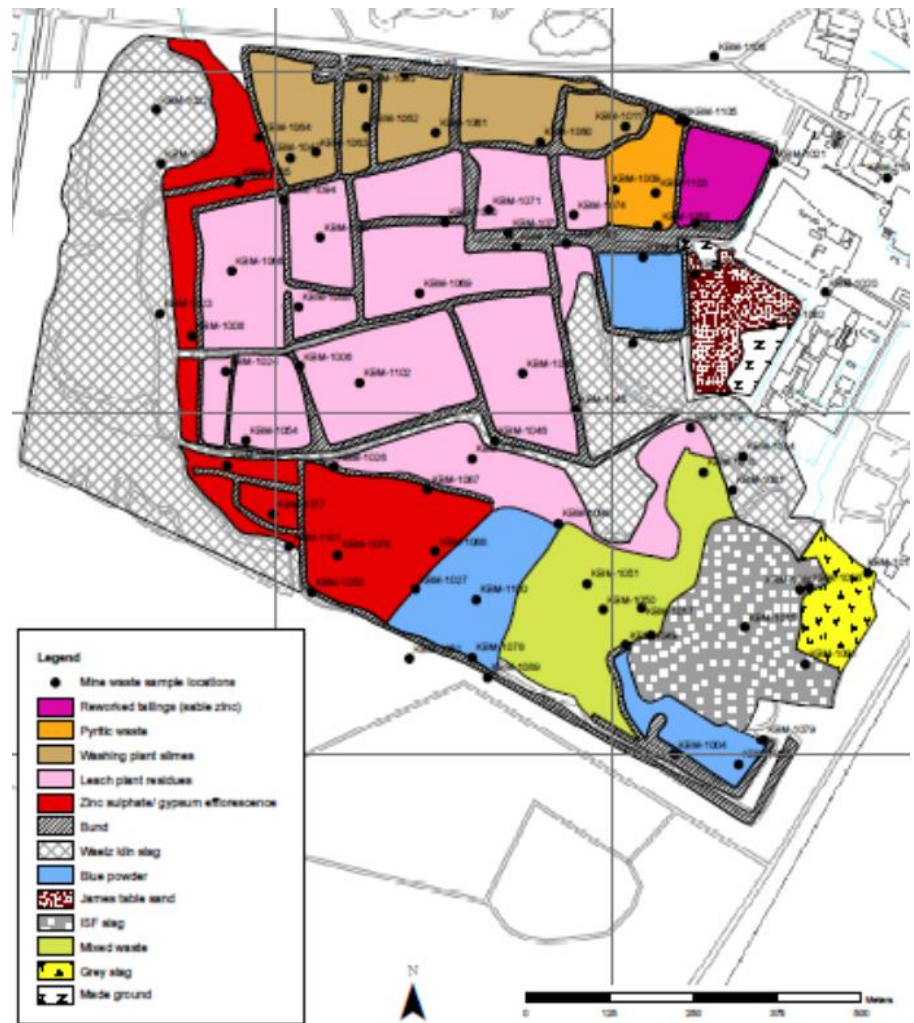
- Black

Site B

- Dark brown




Site C

- Light brown



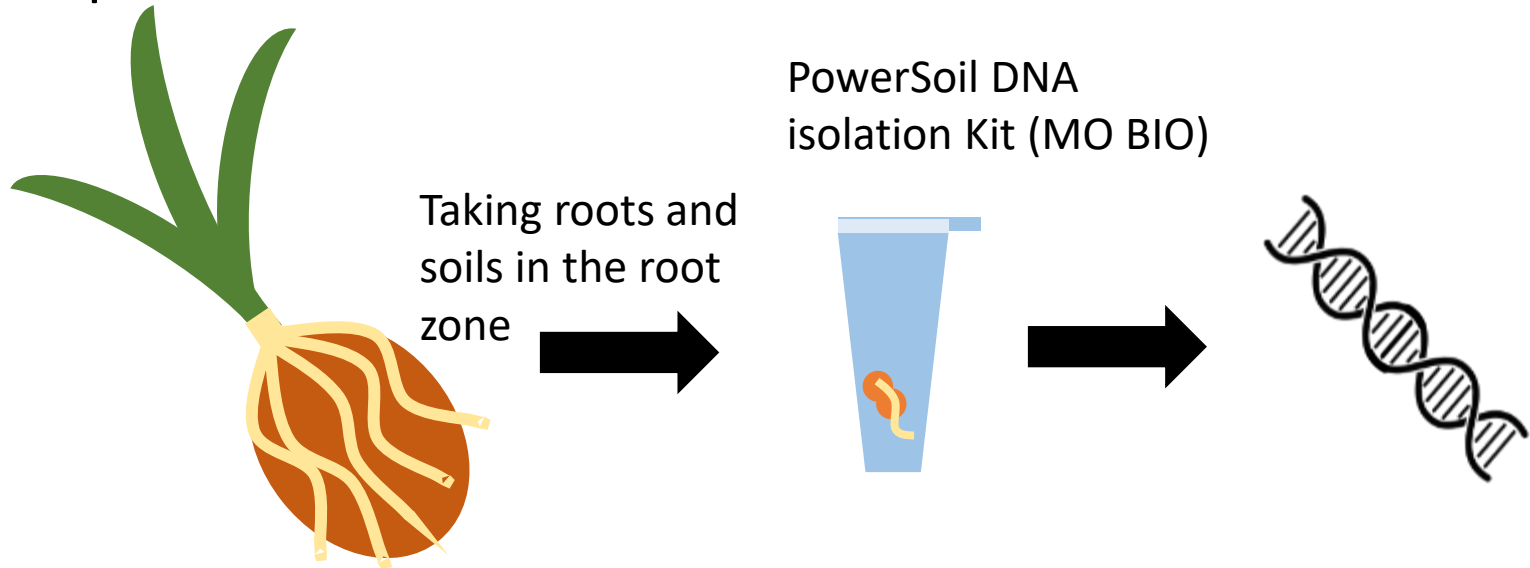
Reference: Kabwe scoping and design study phase 1 complement report by Water Management Consultants Ltd, UK (2006)

Soil properties

	Site A	Site B	Site C
			
pH(H ₂ O)	8.73	6.39	7.22
Total C [g kg dry soil]	148	50.8	9.43
Total N [g kg dry soil]	5.30	3.20	0.38
Total Pb [mg kg dry soil]	3732	11573	19319

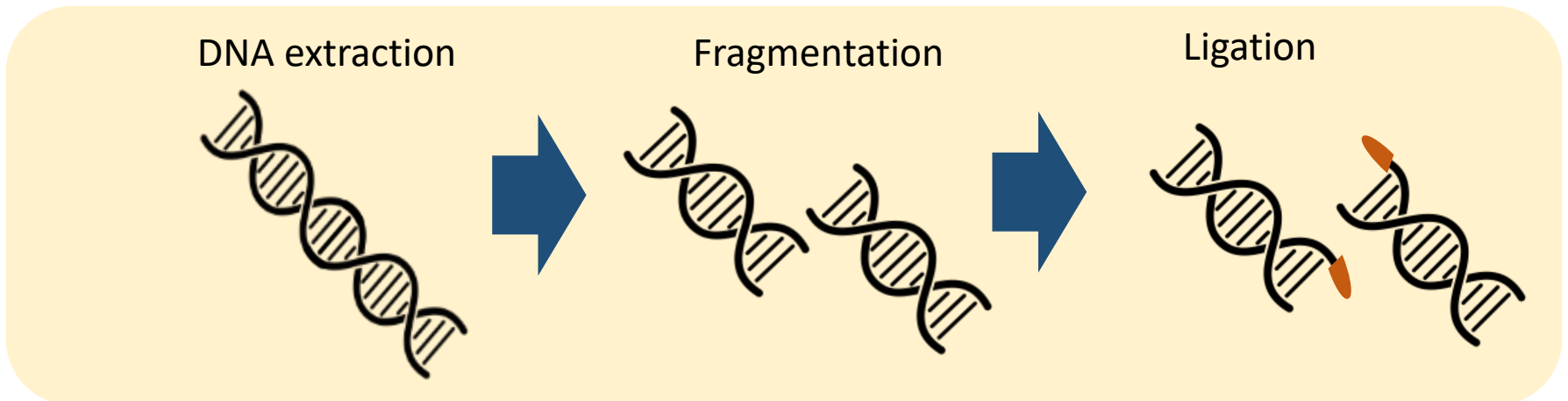
Microbial analysis

- Step 1: DNA extraction



- Step 2: PCR-free preparation

Rapid Barcoding Kit (Oxford Nanopore)

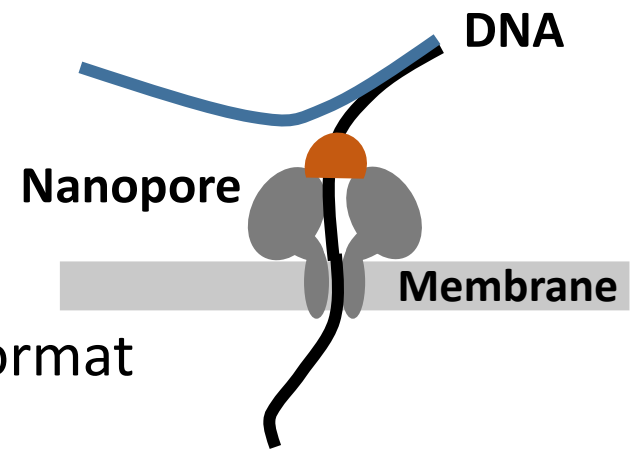


Step 3: Sequencing

- MinKNOW software

- To collect sequencing dataset in Fast5 format
 - Signal data
 - Sequencing reads (e.g. TAACG...)
 - Quality scores (e.g. 19,20,22,18,20)

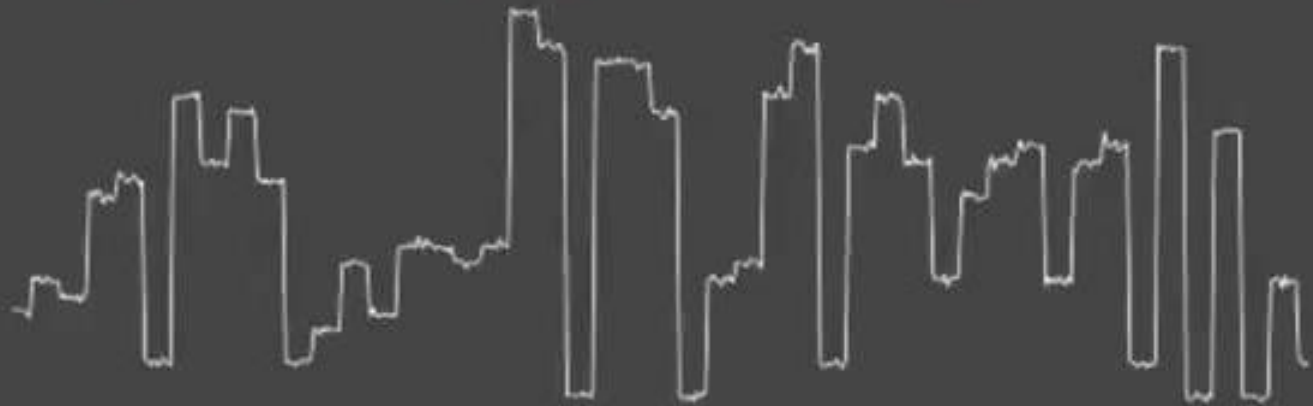
**It shows reliability of the sequencing read data.*



Sequencing reads

3GGCAATAACGTTTATGTTGGTTTCATGGTTTGGTCTAACTTACC

Signal



DATA FOR ILLUSTRATIVE PURPOSES ONLY

Step 4: Sequence classification

- What's in My Pot (WIMP) workflow

1) Filtering

To remove sequences with low quality (QS)
[Averaged QS < 10]

Sample 1

G C C C A T T ...
15, 13, 16, 7, 10, 13, 10...

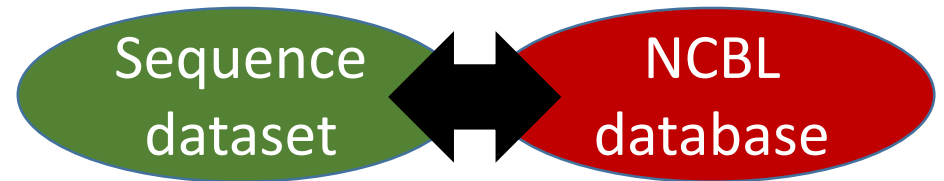
Further analyzed

Sample 2

G C C C A T T ...
5, 9, 10, 7, 10, 9, 10...

Removed

2) Classification

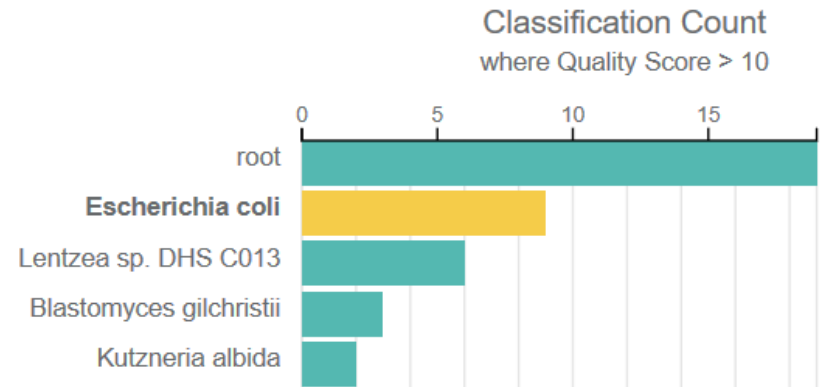


Sample 1

GGTCATTAAATA...

Escherichia coli

GGTCATTAAATA...



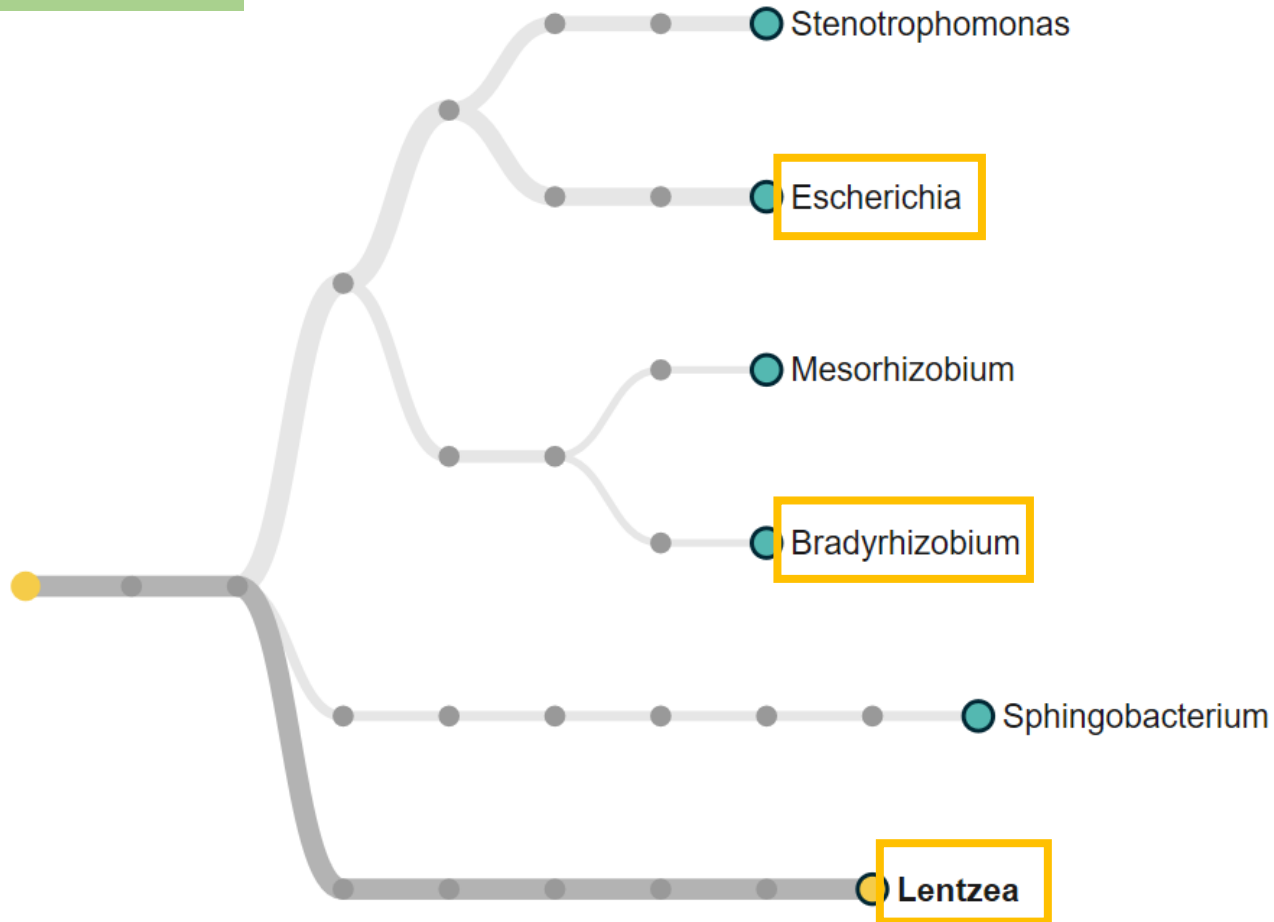
Result and discussion: Classification

	A	B	C
No. of reads	897	220	618
No. of classified reads	106	32	86
No. of identified species	6	8	11
Taxa (QS>10)	Root	Root	<i>Lentzea</i>
	<i>Escherichia</i>	<i>Escherichia</i>	Root
	<i>Lentzea</i>	<i>Bradyrhizobium</i>	<i>Bradyrhizobium</i>
	<i>Bradyrhizobium</i>	<i>Mesorhizobium</i>	<i>Escherichia</i>

- The reads with low quality were removed during the filtering step for the classified reads (Average quality scores 7.15).
- Long reads with higher quality are more likely to be classified (min 5 bases, max 220,480 bases).

Result and discussion: Phylogenetic tree

Site C



Result and discussion: Common bacteria among the different sites

***Escherichia* spp.**

- ❖ Ubiquitous soil bacteria

***Lentzea* spp.**

- ❖ Capable of degrading high molecular weight ester [3]



***Bradyrhizobium* spp.**

- ❖ Symbiotic bacteria of legume plants
- ❖ N₂-fixation [4]
- ❖ Common in South Africa and Western Australia soil [5]

Photo from "A Comprehensive Survey of International Soybean Research - Genetics, Physiology, Agronomy and Nitrogen Relationships"

Conclusions

1. Despite the low quality scores, the MinION data showed the key bacteria in the root zones.
2. *Bradyrhizobium. spp* and *Lentzea. spp* were common bacteria among the three sites.

Future research

- The number of classified sequences should be improved by modifying the protocols.
- Can *Bradyrhizobium. spp* and *Lentzea. spp* detoxify heavy metals?

Acknowledgements

Prof. Ishizuka, Mayumi

Dr Nakayama, Shota

Ms Hirano, Nagisa

Mr Nakata, Hokuto

Mr Muntali, Kabenuka

Mr Mwansa, Mukuka

Ms Saito, Anna

Kampai project

Reference

- Mendes, R., Garbeva, P., and Raaijmakers, J.M. “The rhizosphere microbiome: significance of plant beneficial, plant pathogenic, and human pathogenic microorganisms”, FEMS Microbiology Reviews, Vol. 37, 2013, pp. 634–663.
- Jarerat, A., Pranamuda, H., and Tokiwa, Y. “Poly(L-lactide)-degrading activity in various actinomycetes”, Macromolecular Bioscience, Vol. 2, 2002, pp. 420-428.
- Zahran, H.H. “Rhizobium-Legume Symbiosis and Nitrogen Fixation under Severe Conditions and in an Arid Climate”, Microbiol. Mol. Biol. Rev., Vol. 63, 1999, pp. 968-989.
- Stępkowski, T., Moulin, L., Krzyżańska, A., McInnes, A., Law, I.J., and Howieson, J. “European Origin of *Bradyrhizobium* Populations Infecting Lupins and Serradella in Soils of Western Australia and South Africa”, Appl. Environ. Microbiol., Vol. 71, 2005, pp. 7041-7052
- Board, J.E. (Ed.) "A Comprehensive Survey of International Soybean Research - Genetics, Physiology, Agronomy and Nitrogen Relationships“, 2013